SEROLOGIC SURVEY OF HUMANS FOR *FLAVIVIRUS* WEST NILE IN SOUTHERN MORAVIA (CZECH REPUBLIC)

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SUMMARY

A serosurvey for West Nile virus (WNV) was carried out in 525 persons, using a plaque-reduction neutralization microtest (PRNµT) with Vero cells and Egyptian topotype Eg-101 strain as test virus. The blood sera were collected in four South-Moravian districts (Hodonín 44 persons, Břeclav 102 persons, Znojmo 170 persons, Jihlava 209 persons) of the Czech Republic in the years 1988 and 1989, and stored at –20°C. Antibodies to WNV were detected in only three humans (0.6% seropositivity): one person each in the districts of Hodonín (2.3% persons positive), Břeclav (1.0% positive) and Jihlava (0.5% positive), with the titres of 1:64, 1:32, and 1:32, respectively. All the three sera were negative for antibody to tick-borne encephalitis virus. The results indicate that activity of WNV in southern Moravia was very low before 1990.

Key words: West Nile virus, Flavivirus, Moravia, Czech Republic, serosurvey

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INTRODUCTION

The mosquito-borne West Nile virus (WNV, family *Flaviviridae*) is the aetiologic agent of West Nile fever (WNF). The most serious manifestation of WNV infection is fatal encephalitis and encephalomyelitis in humans and horses, respectively, as well as mortality in certain domestic and wild birds. WNV is transmitted to humans through the bite of an infected mosquito, usually of the genus *Culex* but sometimes also by other mosquito genera. The virus can occasionally be transmitted from person to person through blood transfusion or organ transplantation. WNV occurs in Africa, Eurasia, Australia (as 'Kunjin' virus) and, since 1999, it has also been reported in the Americas.

In the Czech Republic, WNV was isolated from *Culex pipiens* mosquito in the district of Břeclav, southern Moravia, after the big flood in 1997; specific antibodies against this virus were detected in that year in 2.1% of local population, and five cases of WNF were recorded (1–3).

The aim of the present study was to evaluate the activity of WNV in southern Moravia in the past – prior to the flood year 1997, using an indirect method of serological survey of archived human sera.

MATERIALS AND METHODS

Blood Samples

The serological survey was organized by the National Institute of Public Health, Praha and Regional/District Hygienic

Services. Blood samples were collected by physicians (general practitioners) from at random selected healthy individuals, i.e. from those not suffering from fever or any signs of immunodeficiency (age ranged from 19 and 64 in both sexes approximately at the same proportion), in the years 1988 and 1989. The blood samples were centrifuged in laboratory 3 to 6 hours after collection, and the separated sera were stored at –20°C in the Serum Bank of Centre of Epidemiology and Microbiology, NIPH until use. Every blood sample taken was coded according to region, district, physician and patient. Every person participating in the study has provided his/her written consent.

Cell Cultures

Vero E6 and SPEV (pig embryo kidney) cells were serially propagated in Leibowitz L-15 medium (Sigma) supplemented with 10% of foetal calf serum (FCS, Gibco Bio-Cult) and antibiotics.

Viruses

Two viruses were used for PRNT: 1. WNV Eg-101 Egyptian topotype of WNV, lineage 1, passaged 16 times in suckling mouse brain and homogenized in PBS with 0.4% of bovine serum albumin fraction V Sigma). 2. Tick-borne encephalitis virus (TBEV) strain Hypr, Central European topotype of TBEV, passaged 10 times in mouse brain, then 55 times in HeLa cells, and once in suckling mouse brain; homogenized in PBS with 0.4% of bovine serum albumin.

Neutralization Test

Plaque-reduction neutralization test (PRNT) was originally proposed by Madrid and Porterfield (4, 5), and adopted to a microtechnique (PRNµT) on 96-well (flat-bottomed) sterile microplates (Sarstedt) for cell cultures (6). Tested sera were inactivated at 56°C for 30 min, and for screening were diluted 1:4 in L-15 medium; 30 µl of the serum (in duplicate) was mixed in a microplate well with 30-µl test dose of the virus (containing about 50 PFU of WNV, passaged in suckling mouse brain homogenized in PBS with 0.4% of bovine serum albumin fraction V, and centrifuged) in L-15 supplemented with 3% of inactivated FCS, and incubated at 37°C for 60 min. Vero cell suspension (in L-15 with 3% FCS) was then added to each test well (60 µl with 20,000 to 30,000 cells per well), and after an incubation at 37°C for 4 h, 120 µl of carboxymethylcellulose sodium salt overlay (1.5% CMC of medium viscosity BDH in PBS mixed with the same volume of L-15 with 3% of inactivated FCS) was added in each well. Controls included the virus test dose and its titration, immune WNV reference serum; control negative serum; and cells without virus. The microplates, sealed in small polyethylene bags, were incubated at 37°C for 5 days and the cultures then stained with 0.1% acidic solution of naphthalene black (Fluka). Sera reactive with virus, revealing 80% or greater reduction in the number of plaques at the 1:4 dilution at screening (corresponding to the 1:8 final dilution of the serum – after mixing with the virus test dose), were titrated in duplicate by twofold dilutions, and those dilutions corresponding to 80% reduction of plaque numbers were regarded as the antibody titres (PRN μ T_{so}). Reciprocal titres \geq 16 were considered positive. The foetal calf serum used in PRNμT was tested for antibodies against WNV in a separate assay, with negative response.

The positive sera were also tested analogically in PRN μ T against TBEV using SPEV (pig embryo kidney) cells, in order to check for cross reactions with this related *Flavivirus* occurring in Central Europe.

RESULTS

Only three of 525 (0.6%) examined persons reacted with WNV at the titre 1:16 or higher (Table 1): 1/44 (2.3%) persons (no. positive/no. examined) from the district of Hodonín, 1/102 (1.0%) from the Břeclav district, 1/209 (0.5%) from the Jihlava district, but none of 170 persons tested from the Znojmo district. All three WNV-seroreacting persons were also examined in

Table 1. Subjects with specific antibodies against West Nile virus

Code number	Age (years)	Sex	Date of collection	District	PRNµT ₈₀ titre*
3220	46	М	Nov. 1988	Hodonín	1:64
3469	53	M	May 1989	Jihlava	1:32
3720	55	F	March 1988	Břeclav	1:32

^{*} the serum dilution corresponding to 80% reduction of plaque numbers was regarded as the antibody titre

 $PRN\mu T_{80}$ with TBEV, and none reacted with the latter virus at titres 1:16 or higher.

DISCUSSION

The plaque-reduction neutralization test is regarded as the 'gold standard' in *Flavivirus* serology and used for verification of other serological tests (such as ELISA, hemagglutination-inhibition test – HIT) because it is generally more specific and discriminatory. However, it is well known that flaviviruses present a high degree of serological cross-reactivity, even in the neutralization test (5, 7–10). Often several antigenically closely related flaviviruses of the same antigenic group co-occur in one area, e.g., in Central Europe TBEV together with WNV. It is therefore sometimes very difficult to decide which particular virus was responsible for the antibody production.

In this study, we used neutralization with the standard topotype Egyptian strain Eg-101 of WNV and with carefully stored and thermally inactivated serum samples devoid of heparin, citrate, EDTA or any stabilizing substances like merthiolate. We estimated the results conservatively, as the 80% reduction in the number of plaques as an antibody titre cut-off point.

Evidence of the WNV presence in southern Moravia has been based on the virus isolation, five human cases of WNF in 1997, and confirmed by the presence of antibodies in human population that year (1–3). WNV was also isolated previously in neighbouring West Slovakia (11). It was shown later that the WNV strain isolated in South Moravia presents a new, third genomic lineage of the WNV, called 'Rabensburg' (12).

In Belarus areas where WNV strains were isolated from mosquitoes, a bird, and a febrile human patient, the virus antibodies prevailed in 1.7% of the human population (13). In a similar situation, antibodies neutralizing WNV were detected in 2.1% of 619 hospitalized patients or subjects seeking outpatient clinics in southern Moravia in 1997 (3).

CONCLUSIONS

In general, data of the present study indicate indirectly a very limited, if any, WNV activity in southern Moravia prior to 1990, when only 0.6% of persons were seropositive against WNV. Moreover, it could not be excluded that these seropositive persons acquired the WNV infection elsewhere than in southern Moravia; their anamnestic data on travelling abroad could not be ascertained.

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